TR0450 | ACCESSION NUMBER | 76-1521

DDF A:1:08

DATA DOCUMENTATION FORM

TR0450

NOAA FORM 24-13
(4-72)

U.S. DEPARTMENT OF COMMERCE
NATIONAL OCEANIC AND ATMOSPHERIC ADMINISTRATION
NATIONAL OCEANOGRAPHIC DATA CENTER
RECORDS SECTION
ROCKVILLE, MARYLAND 20852

FORM APPROVED
O.M.B. No. 41-R2051

This form should accompany all data submissions to NODC. Section A, Originator Identification, must be completed when the data are submitted. It is highly desirable for NODC to also receive the remaining pertinent information at that time. This may be most easily accomplished by attaching reports, publications, or manuscripts which are readily available describing data collection, analysis, and format specifics. Readable, handwritten submissions are acceptable in all cases. All data shipments should be sent to the above address.

A. ORIGINATOR IDENTIFICATION

THIS SECTION MUST BE COMPLETED BY DONOR FOR ALL DATA TRANSMITTALS

1. NAME AND ADDRESS OF INSTITUTION, LABORATORY, OR ACTIVITY WITH WHICH SUBMITTED DATA ARE ASSOCIATED

Dr. R.T. Cooney
Institute of Marine Sci.
U. Alaska
Fairbanks, Alaska 99701

2. EXPEDITION, PROJECT, OR PROGRAM DURING WHICH DATA WERE COLLECTED

NOAA/BLM/OCS OCSEAP
Zooplankton RU # 156

3. CRUISE NUMBER(S) USED BY ORIGINATOR TO IDENTIFY DATA IN THIS SHIPMENT

Discoverer 810
File ID RTCZ02

4. PLATFORM NAME(S)	5. PLATFORM TYPE(S) (E.G., SHIP, BUOY, ETC.)	6. PLATFORM AND OPERATOR NATIONALITY(IES)		7. DATES	
		PLATFORM	OPERATOR	FROM: MO/DAY/YR	TO: MO/DAY/YR
Discoverer	Ship	USA	USA	8/15/75	8/26/75

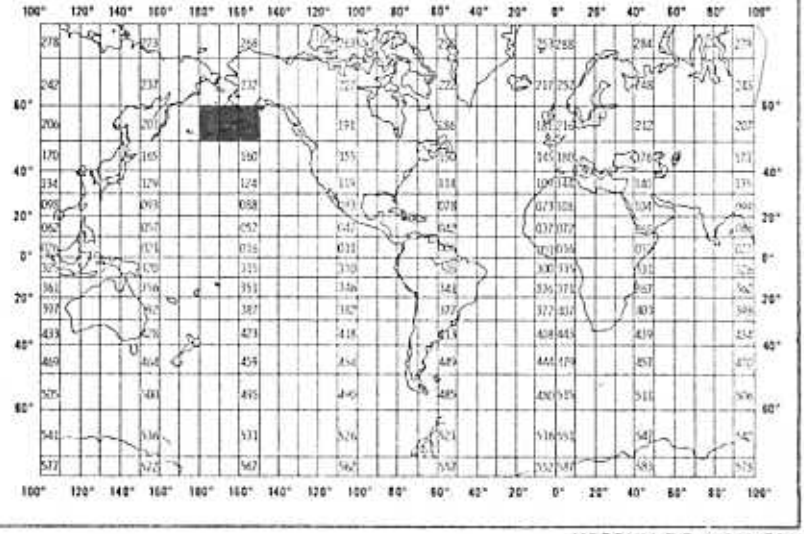
8. ARE DATA PROPRIETARY?

[X] NO [ ] YES

IF YES, WHEN CAN THEY BE RELEASED FOR GENERAL USE? YEAR____ MONTH____

11. PLEASE DARKEN ALL MARSDEN SQUARES IN WHICH ANY DATA CONTAINED IN YOUR SUBMISSION WERE COLLECTED.

GENERAL AREA



9. ARE DATA DECLARED NATIONAL PROGRAM (DNP)?
(I.E., SHOULD THEY BE INCLUDED IN WORLD DATA CENTERS HOLDINGS FOR INTERNATIONAL EXCHANGE?)

[X] NO [ ] YES [ ] PART (SPECIFY BELOW)

10. PERSON TO WHOM INQUIRIES CONCERNING DATA SHOULD BE ADDRESSED WITH TELEPHONE NUMBER (AND ADDRESS IF OTHER THAN IN ITEM-1)

Dr. R.T. Cooney
P.S. Hadley

B. SCIENTIFIC CONTENT

NAME OF DATA FIELD	REPORTING UNITS OR CODE	METHODS OF OBSERVATION AND INSTRUMENTS USED (SPECIFY TYPE AND MODEL)	ANALYTICAL METHODS (INCLUDING MODIFICATIONS) AND LABORATORY PROCEDURES	DATA PROCESSING TECHNIQUES WITH FILTERING AND AVERAGING
Zooplankton Species	Taxon Code # cells/sub sample	1 meter nets 2 meter Tucker Trawls	See procedures enclosed →	

Zooplankton

C. DATA FORMAT

COMPLETE THIS SECTION FOR PUNCHED CARDS OR TAPE, MAGNETIC TAPE, OR DISC SUBMISSIONS.

1. LIST RECORD TYPES CONTAINED IN THE TRANSMITTAL OF YOUR FILE
GIVE METHOD OF IDENTIFYING EACH RECORD TYPE

Record Type I
Record Type II
Record Type III
Record Type IV
Record Type V

2. GIVE BRIEF DESCRIPTION OF FILE ORGANIZATION

Record type I, Record types II, III, IV, V, for 1 meter nets, Record types II, III, IV, V, for 2 meter Tucker Trawl.

3. ATTRIBUTES AS EXPRESSED IN ☐ PL-1 ☐ ALGOL ☐ COBOL ☒ FORTRAN ☐ _____ LANGUAGE

4. RESPONSIBLE COMPUTER SPECIALIST:
NAME AND PHONE NUMBER Cydney Hansen (907) 479-7836
ADDRESS Institute of Marine Science, University of Alaska, Fairbanks, AK 99701

COMPLETE THIS SECTION IF DATA ARE ON MAGNETIC TAPE

5. RECORDING MODE
☐ BCD ☐ BINARY
☐ ASCII ☒ EBCDIC
☐ _____

6. NUMBER OF TRACKS (CHANNELS)
☐ SEVEN
☒ NINE
☐ _____

7. PARITY
☒ ODD
☐ EVEN

8. DENSITY
☐ 200 BPI ☐ 1600 BPI
☐ 556 BPI
☒ 800 BPI
☐ _____

9. LENGTH OF INTER-RECORD GAP (IF KNOWN) ☐ 3/4 INCH ☒ 0.5-0.6 Inch

10. END OF FILE MARK ☐ OCTAL 17 ☒ Octal 23

11. ~~TYPE~~ ~~CONS~~
156, 164 024 RTZO2
Discoverer 810
75/8/9 - 75/8/28 R.T. Cooney
9Trk, 800 BPI,- EBCDIC, No label, ODD

12. PHYSICAL BLOCK LENGTH IN BYTES
80 BYTES

13. LENGTH OF BYTES IN BITS
8 BITS/BYTE

RECORD FORMAT DESCRIPTION

RECORD NAME _____

14. FIELD NAME	15. POSITION FROM - 1 MEASURED IN _____ (e.g., bits, bytes)	16. LENGTH		17. ATTRIBUTES	18. USE AND MEANING
		NUMBER	UNITS		
as per	file Type	'024'		approved 76/05/06	

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USCOMM-DC 44289-P72

D. INSTRUMENT CALIBRATION

This calibration information will be utilized by NOAA's National Oceanographic Instrumentation Center in their efforts to develop calibration standards for voluntary acceptance by the oceanographic community. Identify the instruments used by your organization to obtain the scientific content of the DDF (i.e., STD, temperature and pressure sensors, salinometers, oxygen meters, velocimeters, etc.) and furnish the calibration data requested by completing and/or checking ("✓") the appropriate spaces. Add the interval time (i.e., 3 months, 6 months, 9 months, etc.) if the fixed interval calibration cycle is checked.

INSTRUMENT TYPE (MFR., MODEL NO.)	DATE OF LAST CALIBRATION	INSTRUMENT WAS CALIBRATED BY		CHECK ONE: INSTRUMENT IS CALIBRATED					INSTRUMENT IS NOT CALIBRATED
		YOUR ORGANIZATION (√)	OTHER ORGANIZATION (GIVE NAME)	AT FIXED INTERVALS (√)	BEFORE OR AFTER USE (√)	BEFORE AND AFTER USE (√)	ONLY AFTER REPAIR (√)	ONLY WHEN NEW (√)	(√)
See procedures enclosed.									

NOAA FORM 24-13

USCOMM-DC 44289-P72

EBCDIC - Zooplankton

Octal dump - First 10 records

WR187 03 08/05/76 UTILITY REPORT 750804 PAGE 1

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$ FFILE IN.PHYREC
$ FUTIL IN,,REW/IN/.DUMP/10P/
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BLKs	MODE	CC	WRD#						DENS 800
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			6	036076170760	757727607617	037175772760	761713702004	010020040302		3+"\7+=\G+"\3Z=\G+"\#Y 410 432
			11	613547116534	350065541742	200401002004	010020040100	662403432014		/(P9V)(0V*\K 410 410 410WD3L @
			16	172665552705	720401002004	010020040000				\FV)G5-410 410 400
2		5	1	741713646636	170372370362	745703607417	037075372361	743703606537		%\#UW<\3-\3S%'3+%\3Y=\C/%\3+V\
			6	076676171371	741733467577	276076170765	743717637404	010020040100		7W"\#Z%\.O=\G+"\7V%\\T%410 410
			11	741703607417	036274170100	200401002004	010020040100	200401002004		%\3+%\3S%\10 410 410 410 410 4
			16	010020040100	200401002004	010020040000				10 410 410 410 400
3		5	1	741713646636	170372370362	745703607417	176575372365	761707626537		%\#UW<\3-\3S%'3+%\\V=\CV"\7SV\
			6	076676371363	741703467577	276076170771	743727607404	010020040100		7W"\#T%\3O=\G+"\7Z%\G+%410 410
			11	741703607417	036574170100	200401002004	010020040100	200401002004		%\3+%\3V%\10 410 410 410 410 4
			16	010020040100	200401002004	010020040000				10 410 410 410 400
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			11	741703607417	036274170100	200401002004	010020040100	200401002004		%\3+%\3S%\10 410 410 410 410 4
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			11	741703607417	036274170100	200401002004	010020040100	200401002004		%\3+%\3S%\10 410 410 410 410 4
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			6	076774170361	743743467577	276076170771	745703617564	010020040100		7X%\3/%\LO=\G+"\7Z%'3/=U10 410
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			16	010020040100	200401002004	010020040000				10 410 410 410 400
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			16	010020040100	200401002004	010020040000				10 410 410 410 400
8		5	1	741713646636	170372370362	745703607417	176775372763	747713646537		%\#UW<\3-\3S%'3+%\\X=\GT%\#UV\
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 16  010020040100 200401002004 010020040000
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%\3+%\3V%\10 410 410 410 410 4
10 410 410 410 400
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U.S. DEPARTMENT OF COMMERCE
National Oceanic and Atmospheric Administration
ENVIRONMENTAL RESEARCH LABORATORIES

OUTER CONTINENTAL SHELF ENERGY PROGRAM
JUNEAU PROJECT OFFICE
P. O. BOX 1808
JUNEAU, ALASKA 99802

Date: August 17, 1976

To: Jim Audet
EDS Data Coordinator

From: F. M. Cava FMCava
Assistant Data Manager
Juneau Project Office

Subj: Data Submission for R.U. 156, 164, 426

Under separate cover one magnetic tape and DDF is being sent to you. The tape is labelled as follows:

156, 164 024 RTZ02
Discoverer 810
75/8/9 - 75/8/28 R.T. Cooney
9 TRK, 800 BPI, EBCDIC, No Label, Odd

cc: R. Cooney
R. Hadley
D. Day

76-1521

PROCEDURES AND QUALITY CONTROL

for

ZOOPLANKTON AND MICRONEKTON STUDIES IN THE BERING - CHUKCHI/BEAUFORT SEAS

as used by:

R. T. Cooney, Principal Investigator
Contract Number 03-5-022-56
Task Order #13, R.U. #156/164

FIELD PROCEDURES

1. Meter Net

A 1-m (dia) net of 0.333-mm Nitex was fished vertically at selected stations. The net was lowered backwards through the water column to within 5 m of the sea bed (at locations 200-m or shallower) or generally 200-m below the surface, (certain casts were taken as deep as 500 m below the surface), and then retrieved at about 1 m/sec. A flow meter was not used to measure volume filtered, nor was the wire angle measured. The net was rinsed thoroughly at the surface and the catch preserved in 5-10% formalin-seawater. The sample was labled by station name, cruise, date, time of day, depth of tow, and gear type. Samples were returned to the University of Alaska Marine Sorting Center for processing.

2. Tucker (NIO) Trawl

A 2-m (square mouth) Tucker trawl of 1/8 inch knotless nylon was fished in an open double oblique mode at selected stations. The trawl was lowered and retrieved with the vessel underway at 3-6 m/sec. For most tows, the maximum depth fished was measured using a bathykymograph, and the volume filter obtained from a flow meter in the mouth of the net. The trawl was cleaned after each haul and the catch preserved in 5-10% formalin-seawater. The sample was labeled by location, cruise, time of day, maximum depth, and gear type. Catches were returned to the University of Alaska Marine Sorting Center for processing.

3. Bongo Net

60-cm bongo net systems were used (on rare occasions) with Nitex .333-mm mesh netting to sample zooplankton and micronekton. The net was placed in the water with the vessel underway @ 2-3 knots and fished in an open double oblique mode between the surface and 10-m above the bottom at depths shallower than 200-m, and between the surface and 200-m depths at deeper locations. The net was set and retrieved at approximately 60-m per minute.

On deck, the catch was rinsed from the netting into collecting cups and transferred to sample bottles for processing at the Marine Sorting Center. Unless the catch was large, organisms from both nets were poured together and fixed in 10% formalin-sea water. The amount of water filtered during a tow was measured using a General Oceanic Flowmeter mounted in the mouth of one of the paired nets.

LABORATORY AND SORTING PROCEDURE

All incoming samples are checked for proper preservation and inventoried.

Tucker Trawl

1. Formalin is rinsed from sample.

2. Sample is split, using plankton splitter, until the most abundant species numbers between 50-100 individuals. Each

successively small fraction (i.e., 1/2, 1/4, 1/8, etc.) is placed in two separate bottles.

3. The most abundant species (in the smallest sample fraction) is innumerated and identified.

4. Each successively larger sample portion is analyzed for the less abundant species. When 50-100 specimens of any given species have been sorted out, or when the sample fraction is so concentrated that the specimens cannot be sorted quantitatively, then the number of specimens picked out and the sample fraction from which they were picked is recorded. The process is continued until each species is recorded, along with the sample fraction analyzed for that species and the number of individuals counted.

5. Examples of unusual species or abnormal specimens are saved out as vouchers. Vouchers of all the species collected during the OCS project have been saved.

6. After analysis the samples are recombined, put in a scotch brand plastic bag, sealed, and stored in pasteboard boxes. Lists are kept of all samples archieved in this way.

1-m Net/Bongo Net

1. Formalin is rinsed from the sample.

2. Sample is split in half, one of the halves is quartered.

3. One-fourth of sample is put in a jar for dry weight analysis.

4. One-fourth of the sample is archieved in plastic bags as described above.

5. The microneckton is picked from one-half of the sample, identified and innumerated. It is then recombined with the sample.

6. Sample diluted to a known volume (200 to 2000 mls.), agitated until organisms are evenly distributed, then sub-sampled with a 5 ml. Stempel pipette.

7. A 5-ml sub-sample is rinsed into a petri dish for counting.

8. One or two additional 5-ml sub-samples are taken as in step #7, without further dilution, agitating each time (if very few organisms are present in sample, two or more 5-ml sub-samples may be placed in the same petri dish).

9. Sub-sample is counted and enumerated on data sheet -- if substantially fewer than 100 of the dominant species were counted additional sub-samples are counted until approximately 100 have been counted.

10. All sub-samples are combined with one-half split of sample and stored.

Dry Weight

1. Weighing pans are placed in drying oven (at 60 degrees C.) for 6-8 hours.

2. They are removed, placed in a dessicator until they come to room temperature, and then weighed to the nearest mg.

3. The sample is rinsed and transferred to the weighing pan.

4. Samples are dryed at 60 degrees C for about 24 hours, weighed, dryed for an additional 24 hours and weighed again.

5. Before weighing, the samples are placed in the dessicator and allowed to cool to room temperature.